A COMPREHENSIVE TRICHOME INVESTIGATION OF *Vitex* L. (LAMIACEAE MARTINOV) IN PENINSULAR MALAYSIA AND ITS TAXONOMIC SIGNIFICANCE

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Abstract. The micromorphology and anatomy of trichomes of the leaves of 11 species of the genus Vitex (V. negundo, V. glabrata, V. gamosepala, V. longisepala, V. millsii, V. pinnata, V. quinata, V. trifolia, V. vestita, Vitex sp.1 and Vitex sp.2), one subspecies (V. trifolia subsp. litoralis) and two varieties (V. negundo var. bicolor and V. negundo var. cannabifolia) were investigated using light and scanning electron microscopy which were collected from Peninsular Malaysia. Data were gained both from herbarium and field study. Two basic types of trichomes were identified; non-glandular and glandular trichomes. The non-glandular trichomes are subdivided into simple and branched trichomes (N₁, N₂, N₃, N₄, N₅, N₆ and N₇) while glandular trichomes are subdivided into two subtypes; stalked and sessile (G₁, G₂, G₃, G₄, G₅ and G₆). Trichome distribution and cover within the species provide valuable characters for comparison hence have taxonomic value. Furthermore, trichomes show considerable variation among the species. The diversity of the characters produced an interspecific variation among the genus members, hence can be used to delineate one species from another. The presence of G₂, N₆ and N₇ types of trichome helps to differentiate the species, therefore verifying them belongs to the same genus hence proving the taxonomic significance. This study implicates that leaf trichome characters can be used in identification of species, subspecies and variety of this genus. The new data might propose some changes to the placement of taxa hence can be devoted as an improvement for the previous study.

Keywords: Vitex sp., trichomes, lamiaceae, leban, taxonomy

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Introduction

Lamiaceae is the largest family in the order Lamiales [1] with 236 genera and 7,173 species which is widely distributed globally [2]. *Vitex* is one of the important genus belongs to the Labiatae [3] which currently group into Lamiaceae [4]. There are 250 known *Vitex* species worldwide with 25 species occurring in Southeast Asia [5], while species in Peninsular Malaysia was first described by Lam [6] with 46 species, followed by Ridley [7] with 15 species, both Kochummen [8] and Turner [9] recorded ten species and Bramely [10] recorded 13 species. The most important characters in *Vitex* species are its simple and palmate leaves [10] arranged oppositely and *Vitex* in Malaya are nearly all trees or shrubs [11].

Trichomes are the common characters in the family Lamiaceae [12] which are widely distributed over the aerial reproductive and vegetative parts of plants of Lamiaceae and are usually identified as glandular and non-glandular trichomes [13,14]. Trichomes are well known in Lamiaceae and other related families such as Verbenaceae and Scrophulariaceae as one of the important characters which has the taxonomic value as well as contributing implication in systematics and phylogenetics [13,15-19]. Furthermore, trichome micromorphology has been suggested to be useful in the phylogeny reconstruction [16,20] and systematics of Lamiaceae [13]. Moreover, there are a number of research on comprehensive trichome micromorphology of Lamiaceae been done all over the world including *Phlomoides* [15], *Leucas* [21], *Salvia* [22] and *Vitex* [23]. However, there are only a few research contributed with detailed analysis of trichomes in *Vitex* especially in Peninsular Malaysia.

To date, a list of taxonomic studies have been conducted on *Vitex* in Peninsular Malaysia have been recorded by many researchers [3,7,8,10,24-27] but none of them comprehensively classify the importance of trichomes in the taxonomy of this genus. Thus, a comprehensive study which includes morphology, anatomy and micromorphology of trichomes in *Vitex* of Peninsular Malaysia is still poorly documented. Therefore, this study focuses not only on the morphological character of trichomes, but also its anatomical and micromorphological characters which proved that it can be used as a taxonomic marker.

Materials and Methods

Trichomes of 11 species of the genus *Vitex* (*V. gamosepala, V. glabrata, V. longisepala, V. millsii, V. negundo, V. pinnata, V. quinata, V. trifolia, V. vestita, Vitex* sp.1 and *Vitex* sp.2), one subspecies (*V. trifolia* subsp. *litoralis*) and two varieties (*V. negundo* var. *bicolor* and *V. negundo* var. *cannabifolia*) were investigated. Herbarium specimens were investigated mainly in Forest Research Institute Malaysia (KEP) and Universiti Kebangsaan Malaysia (UKMB). Specimens were also observed virtually from KEW (K), Singapore Botanic Garden (SING), Royal Botanic Gardens, University of South Florida (USF) and New York Botanical Garden (NY) for the purpose of species identification. A total of 509 specimens of *Vitex* were observed. At least five specimens of each species were analysed to ensure certainty of trichome characters of one species among different specimens. While fresh samples were collected through a series of fieldwork in Peninsular Malaysia. Species were collected at Pasoh Forest Reserve, Negeri Sembilan; Sungai Dara Forest Reserve, Perak; Proton City, Tanjong Malim, Perak; Gunung Lambak Forest Reserve, Johor; Cameron Highland, Pahang; Herb Walk Garden, Langkawi; and Kota Bharu, Kelantan. Voucher

herbarium prepared were deposited at Universiti Pendidikan Sultan Idris herbarium (Voucher number: SMZ001 – SMZ0052).

Herbarium and field samples were studied morphologically by following Veldkamp [28] and a combined method of Corner [3] and Kochummen [8]. The surface of the leaves was examined with an Olympus S2 111 stereo microscope. Full attention was paid to the type, density, shape and size of trichomes. All terminology attributes of the studied species were evaluated based on Hickey [29] and Ellis [30]. While for anatomical study, the dissections of petiole, lamina and midrib of the leaflets were done using a microtechnique procedure based on modification of reported methods by Nor Nafizah [31], Cutler [32] and Johansen [33].

Fresh samples were fixed with Formaldehyde Alcohol Acetic Acid (FAA) solution for 24 hours, while leaves from herbarium specimens were boiled 5 – 10 minutes, followed by fixing in the same solution as fresh leaves. Direct sectioning using a microtome at 2 μm thick was done. Sections were then bleached with Clorox for five minutes followed by gently and thoroughly rinsing with distilled water for a 3 – 5 times before staining in a few drops of Safranin and Alcian Blue. The sample was soaked for 3 – 5 minutes and rinsed in distilled water to clear off color. The sections were then underwent an ascending alcohol series, 50 %, 70 %, 95 % and 100 %, 2 – 5 minutes for each for the purpose of dehydration, then 1:1 Xylene + alcohol for 3 – 5 minutes and then rinsed with pure Xylene. The sample was then mounted onto slides with cover slips by using Canada Balsam and kept in 60 °C. Microimages were captured using Nikon Eclipse 2000-U microscope fitted with image analyzer. Anatomical descriptions were made according to Metcalfe and Chalk [18,34] while anatomical slides prepared were kept at Plant Microtechnique Laboratory, Centre for Biodiversity and Conservation, Faculty of Science and Mathematics, Universiti Pendidikan Sultan Idris for future references.

For the scanning electron microscopy (SEM), samples from the leaves were cut with size 0.3 to 0.5 cm³ using knife and then were mounted directly onto stub by using double sided tape and gold-coated. After that, the samples were examined at different several magnifications and the best were selected normally at 500 to 2 000 magnification to observe the trichome clearly. The SEM micrographs were taken at an accelerating voltage of 10–15 kV. The type of trichome was described and classified. The general classification scheme and the terminology were done according to Metcalfe et. al. [34] and Payne [35].

Results and Discussion

Types of Trichome in Vitex

The basic types of the observed trichomes were illustrated in Figure 1. The characteristic features and their distribution among the studied species were summarized in Table 1. Selected SEM and light microscopy (LM) micrographs of trichome types were presented in Figures 2-5. Morphological, micromorphological and anatomical observations of trichomes of *Vitex* in Peninsular Malaysia shows variation from unicellular to multicellular, conical to elongated, smooth to ridges, curved at apices to blunt and glabrous to very dense. Trichomes were present on both abaxial and adaxial epidermises of the leaves, petiole and stalk of all species examined.

There were 13 types of trichomes identified in this research. Each of them was categorized into two; glandular and non-glandular trichomes, supporting the classification of trichomes in Lamiaceae which was done by Cantino [13]. Based on the variation observed, glandular trichome can be subdivided into two subtypes: stalked (*V. longisepala, V. gamosepala* and *V. trifolia* subsp. *litoralis*) and sessile (*V. millsii, V. negundo, V. vestita, V. negundo* var *cannabifolia, V.* sp.1 and *V.* sp.2). There were two types of stalked glandular trichome observed in this study; G₁ and G₂. Stalked glandular trichome can be short with a capitate head. The heads of the glandular trichome were either bicellular; G₁ (*V. longisepala, V. gamosepala* and *V. trifolia* subsp. *litoralis*) or multicellular, G₂ (*V. vestita, V. negundo, V. millsii, V. negundo* var *cannabifolia, V. negundo* var *bicolor, V.* sp.1 and *V.* sp.2). While nonglandular trichomes were divided into two subtypes: unbranched; N₁, N₂, N₃, N₄ and N₅ and branched trichomes; N₆ and N₇.

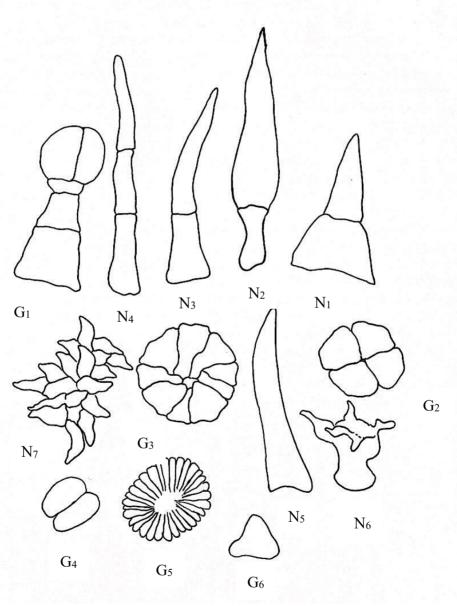


Figure 1: Types of trichome identified in *Vitex* sp. Glandular trichomes; G₁, G₂ (stalked), G₃, G₄, G₅ and G₆ (sessile), Non-glandular trichomes; N₁, N₂, N₃, N₄, N₅ (unbranched), N₆ and N₇ (branched) [18,34]

Table 1: The main types of trichomes presence and some of the characteristics features in investigated *Vitex* species

Species	Surface	Density		Size	Glandular trichome						Non-glanduar trichome						
		Adax	Abax	range	G_1	G_2	G_3	G_4	G_5	G_6	N_1	N_2	N ₃	N_4	N_5	N ₆	N_7
V. pinnata L.	Papillate	Dense	Dense	6 – 44μm	+	-	+	+	+	+	-	-	-	+	+	-	-
V. glabrata R. Br.	Smooth	Glabrous	Sparse	$\begin{array}{c} 43-\\ 80\mu m\end{array}$	-	-	-	-	+	-	+	+	-	-	-	-	-
V. quinata (Lour.)	Smooth	Sparse	Very dense	33 - 114μm	+	-	-	-	+	-	+	+	+	-	-	-	-
V. longisepala King & Gamble	Papillate	Dense	D ense	8 - 117μm	+	-	+	+	-	-	+	-	+	+	+	-	-
V. trifolia L.	Papillate	Dense	Very dense	40 - 85μm	+	-	-	-	-	-	-	+	+	-	-	-	-
V. trifolia subsp. litoralis Steenis	Smooth	Dense	Very dense	35 - 75μm	+	-	-	-	-	-	-	+	+	-	-	-	-
V. gamosepala Griff.	Papillate	Sparse	Sparse	45 - 89μm	-	-	+	-	-	-	+	-	-	+	-	-	-
V. millsii M.R. Hend.	Papillate	Sparse	Sparse	74 - 112μm	+	-	+	-	+	-	+	-	+	+	-	-	-
V. vestita Wall. Ex Schauer	Papillate	Dense	Very dense	38 - 576μm	+	-	+	+	-	-	+	-	+	+	+	+	+
V. negundo L.	Papillate	Sparse	Very dense	18 - 113μm	+	-	-	-	-	-	+	+	+	+	-	-	-
V. negundo var. bicolor (Willd.) H.J. Lam	Papillate	Sparse	Very dense	15 - 110μm	+	-	-	-	-	-	+	+	+	+	-	-	-
V. negundo var. cannabifolia (Siebold & Zucc.) HandMazz.	Papillate	Sparse	Sparse	25 - 109μm	-	+	-	-	-	-	+	+	+	+	-	-	-
V. sp.1	Smooth	Sparse	Dense	70 - 135μm	-	-	+	-	-	-	+	-	+	-	-	-	-
V. sp.2	Smooth	Sparse	Dense	28 - 201µm	+	-	+	-	-	-	+	-	+	-	-	-	-

Trichome Description

The trichome characters which found to have a taxonomic value in this study were presence of glandular trichome and non-glandular, presence of branched trichomes (dendroid or peltate), number of cells (unicellular or multicellular), density of trichomes (abaxial and adaxial surface), size of trichomes (glabrous, sparse, dense or very dense) and surface of the leaves (smooth or papillate). There are 13 types of trichome characters identified in this study (Figure 1) which were described based on [12,13]. All of these trichome types are found to be useful and valuable taxonomically especially in delineating the species, further aid the identification of the species. Type of trichome shows significant variability among different species, but is constant among different populations of one species, and therefore, affords valuable characters in delimitation of sections and species [15].

The trichome varies from a simple unbranched trichome, single cell, into scales and dendritic branched trichomes. The glandular trichome is observed in six different types. The G₁ type has a simple, short, thin capitate (Figure 2(1)). It is an unbranched trichome which is observed in *V. pinnata*, *V. glabrata*, *V. longisepala*, *V. trifolia*, *V. milsii*, *V. vestita*, *V. negundo*, *V. negundo* var. *bicolor* and *V.* sp.2. The G₂ type is a multicellular scale, rounded with four cells divided (Figure 2(2)). This was observed only in *V. negundo* var. *cannabifolia*. Type G₃ is a glandular trichome, multicellular with more than four cells divided (Figure 2(3)). This type of trichome was observed in *V. pinnata*, *V. glabrata*, *V. longisepala*, *V. gamosepala*, *V. millsii*, *V. vestita*, *Vitex* sp.1 and *Vitex* sp.2. Type G₄ is a bicellular scale, oval with two cells divided (Figure 2(4)) observed in *V. pinnata*, *V. longisepala* and *V. vestita*. While type G₃ is subsessile glandular trichome, rounded with more than ten cells (Figure 2(5)) observed in *V. pinnata*, *V. glabrata*, *V. quinata* and *V. millsii*. Type G₆ is a unicellular triangle shaped trichome (Figure 2(6)) which was observed only in *V. pinnata*. The result has supported the findings of [13].

The rest other seven types are non-glandular trichomes. The non-glandular trichome identified in this study varies from a single-celled to multi-celled which is common in Vitex. The N₁ type is an unbranched trichome with short and thin prickles (Figure 2(7) and (8)). This N₁ type is a multicell which was observed in V. glabrata, V. quinata, V. longisepala, V. gamosepala, V. millsii, V. vestita, V. negundo, V. negundo var. cannabifolia, V. negundo var. bicolor, V. sp.1 and V. sp.2. The N₂ type is a simple multicellular unbranched trichome, with a Christmast tree feature (Figure 3(9)) which were observed in V. glabrata, V. quinata, V. trifolia, V. negundo, V. negundo var. bicolor and V. negundo var. cannabifolia. The N₃ type has a simple long multicellular trichome with two cells divided (Figure 3(10)), observed in V. quinata, V. longisepala, V. trifolia, V. millsii, V. vestita, V. negundo, V. negundo var. bicolor, V. negundo var. cannabifolia, V. sp.1 and V. sp.2. Besides that, the N₄ type has long multicellular trichomes with three cells divided (Figure 3(11)). Species with this type of trichome were observed in V. pinnata, V. quinata, V. gamosepala, V. millsii, V. vestita, V. negundo, V. negundo var. bicolor and V. negundo var. cannabifolia. The Ns type has simple, long and thin unicellular trichome (Figure 3(12)) which was observed in V. pinnata, V. longisepala and V. vestita. The N₆ type is a sessile, non-glandular scale with multicelled (Figure 3(13)). This type of trichome was only observed in V. vestita. The N_7 type is a sessile, nonglandular scale with multicelled (Figure 3(14)). This type of trichome was also observed only in V. vestita. Based on the size, sessile glandular trichome type are the shortest (from 5.59 µm) in V. trifolia, while longest are N₄ type which were found in V. vestita (up to 576 µm). Figure 5 and 6 shows the comparison of trichomes between LM and SEM micrographs of trichome types present in Vitex species.

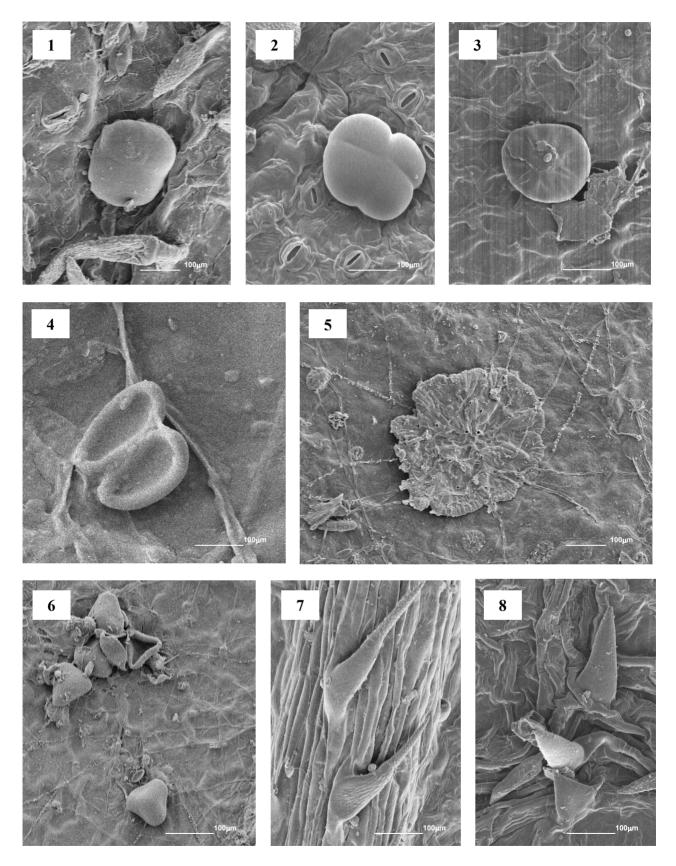


Figure 2: SEM micrographs of *Vitex* sp. in Peninsular Malaysia: (1) *V. quinata*, (2) *V. negundo* var. *cannabifolia*, (3) *V. gamosepala*, (4) *V. longisepala*, (5, 6) *V. pinnata*, (7) *V.* sp. 2 and (8) *V. negundo* var. *cannabifolia*. Scale = 100μm

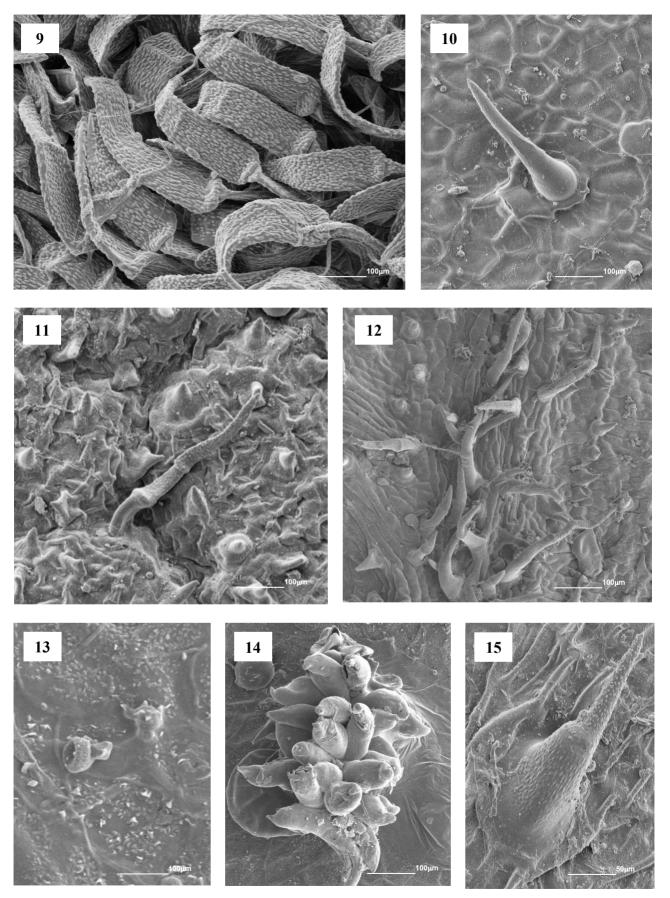


Figure 3: SEM micrographs of *Vitex* sp. in Peninsular Malaysia: (9) *V. vestita*, (10) *V. longisepala*, (11, 12) *V. negundo*, (13, 14) *V. vestita* and (15) *V.* sp. 2. Scale: $(9-14) = 100 \mu m$, $(15) = 50 \mu m$

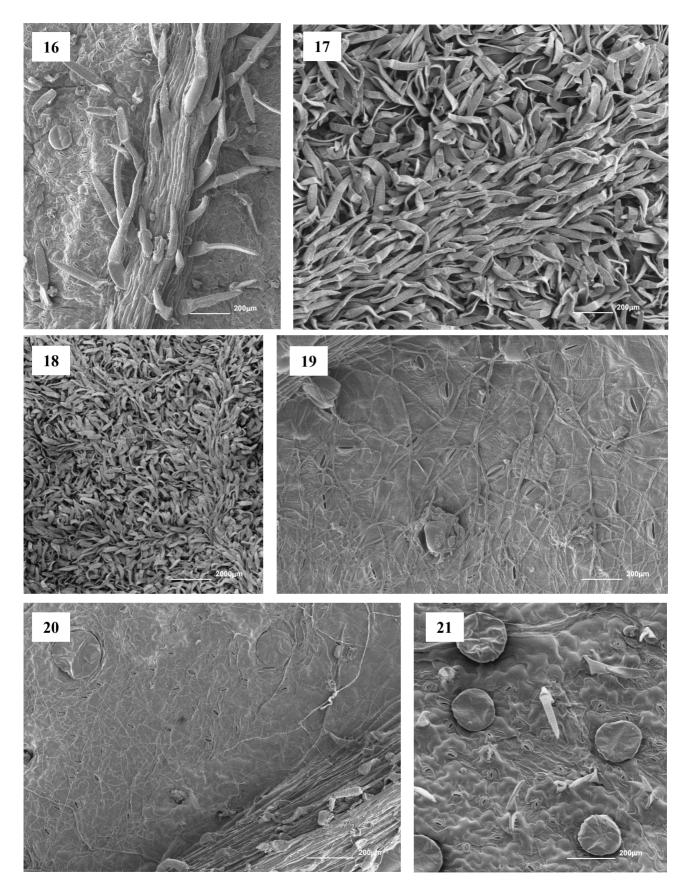


Figure 4: SEM micrographs of *Vitex* sp. in Peninsular Malaysia: (16) *V. negundo* var. *cannabifolia*, (17) *V. negundo*, (18) *V. negundo* var. *bicolor*, (19) *V. pinnata* and (20) *V. glabrata*, (21) *V.* sp.2. Scale = 200μm

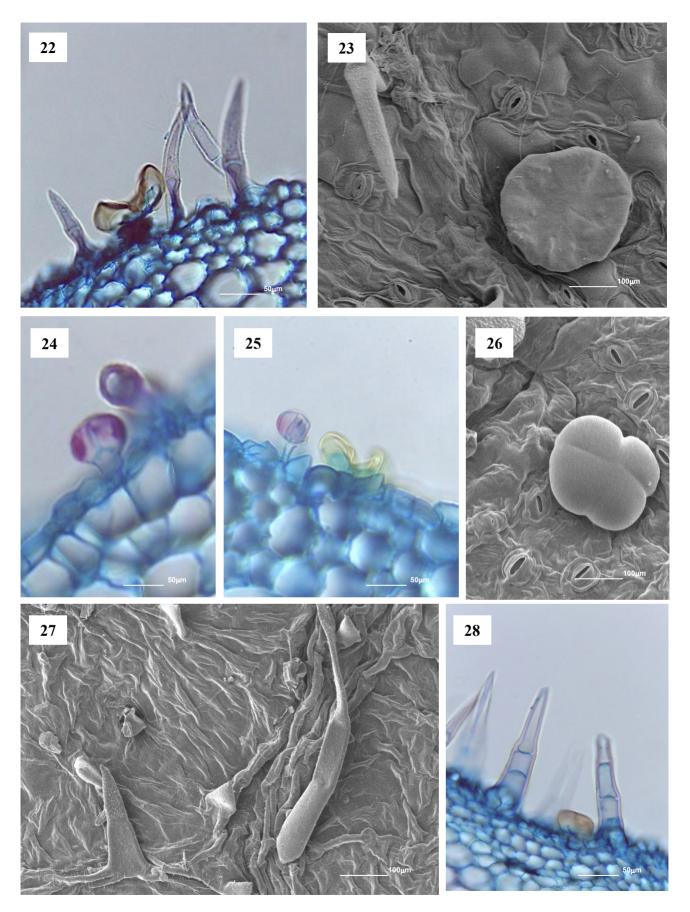


Figure 5: (22, 24, 25, 28) LM micrographs, (23, 26, 27) SEM micrographs of *Vitex* sp. in Peninsular Malaysia (22) *V. millsii*, (23) *V. gamosepala*, (24) *V.* sp. 2, (25) *V.* sp. 1 (26) *V. negundo* var. *cannabifolia*, (27) *V. millsii*. Scale: (22, 24, 25, 28) = 50μm, (23, 26, 27) = 100μm

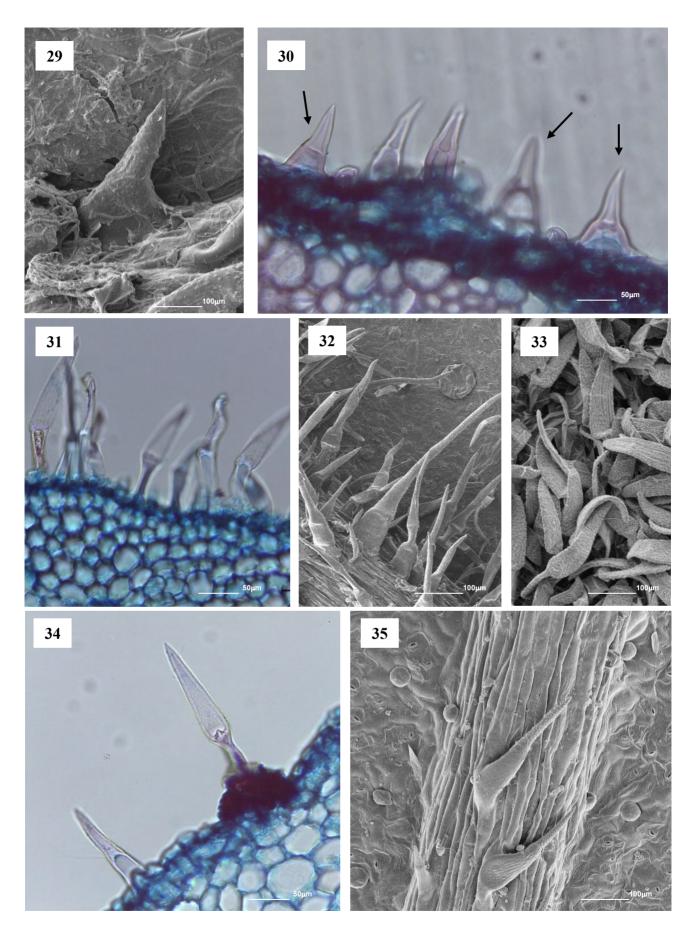


Figure 6: (30, 31, 34) LM micrographs, (29, 32, 33, 35) SEM micrographs of *Vitex* sp. in Peninsular Malaysia, (29, 30) *V. millsii*, (31, 32) *V. negundo* var. *cannabifolia*, (33) *V trifolia* and (34, 35) *V. millsii*. Scale: (22, 24, 25, 28) = 50μm, (23, 26, 27) = 100μm

Diagnostic Characters

The classification of trichomes presented in this study significantly shows the importance of trichome characters within the genus and family. Analysed images obtained support the general classification and description of the trichome in the genus *Vitex* in Peninsular Malaysia which have been updated by Bramely [10]. Furthermore, these findings are consistent with the previous studies in Lamiaceae confirming the importance of trichome characters in taxonomic identification at different infrageneric levels [36]. Moreover, morphological and trichomes distribution also have an important taxonomic significance at species level. The presence of trichome in each of the *Vitex* species proved that trichome characters have important taxonomic value. This has been agreed by Seyedi and Salmaki [15] that trichome morphology is more valuable to separate the species within sections. Furthermore, types of trichome show considerable variability among species in different sections or series, and therefore, affords valuable characters in delineating of sections and species [14].

Glandular trichomes are one of the important taxonomic characters in Lamiaceae [13,37]. Glandular trichomes have taxonomic value at both specific and subspecific and also can be considered as phylogenetically informative characters [20]. For example, the finding in this study shows that the most common types of trichome shared with almost all species of *Vitex* are type G_1 and G_3 . Moreover, [38,39] successfully divided two sections involving Lamiaceae species based on the presence of glandular trichomes. Furthermore, Nor Nafizah et al. [40] agreed that some limited characters are found to be useful in delineating *Vitex* species. Therefore, type G_2 trichome in this study can only be found in *V. negundo* var. *cannabifolia* hence can be used as limited character to delineate the species in the genus.

Besides that, the identification of non-glandular trichomes proved the important of these characters within the genus. Variation in non-glandular trichome characters appears to have specific value, especially to separate different species within the genus [14]. For instance, there are types of trichomes which are N₆ and N₇ only present in *V. vestita* which made these characters to be used in delineating the species. On the other hand, the most common types of trichome characters shared by *Vitex* species are N₁ and N₃. Obviously, this has supported the acceptance of *V. negundo* var. *cannabifolia* as a variety of *V. negundo* L. instead of *V. cannabifolia* before this because of it sharing most common characters, not only morphologically, but anatomically and micromorphologically. While there are two unidentified *Vitex* sp.; *V.* sp.1 and *V.*sp.2 which are found to share most of the common trichome characters shared with other *Vitex* species, hence supporting their belongings in this genus.

On the other hand, trichome distribution on both abaxial and adaxial epidermises within the species provide diagnostic characters for comparison. It was shown that trichomes are generally more commonly distributed on the abaxial than the adaxial surface of the leaves supporting research conducted by Celep et al. [41]. As shown in Table 1, in all species investigated, N₁, N₂, N₃ and N₄ types occur in almost all species and their distribution differs in each of the species. N₁ and N₃ were the most frequent presents. In most species, non-glandular trichomes are mainly found on the abaxial surfaces, especially denser on the leaf margin (Figure 3(16)). In some other species including *V. negundo*, *V. negundo* var. *bicolor*, *V. negundo* var. *cannabifolia*, *V. trifolia*, *V. trifolia* subsp. *litoralis* Steenis and *V. quinata* non-glandular trichomes are distributed over the epidermis as densely as those along the veins (Figure 3(17) and (18)). While *V. glabrata* has the least distribution of trichome on both the abaxial and adaxial surface of the leaves.

Conclusions

Since trichomes were present in plants in a great variety of forms, and are sometimes very complex structurally, the trichome in *Vitex* can be denoted as a diagnostic characteristic from this study. In summary, all the evidence gathered supported that *Vitex* is a genus of Lamiaceae family and therefore, the present study clearly revealed that the diversity of the trichomes in *Vitex* species can be used in identifying the species hence contributing in solving the taxonomic problem. The diversity of the characters produced an interspecific variation among the genus members, hence can be used to delimit one species from another. There are certain trichome characters in detail that are found to be useful in delineating the Vitex species. V. negundo var. cannabifolia for instance is the only species in the members that have the G₂ type of trichome. High similarity in trichome morphological, anatomical and micromorphological characters of all 11 species, one subspecies and two varieties of *Vitex* studied suggest and support that they belong to the same genus. Nonglandular and glandular trichomes are very helpful to differentiate the species. Hopefully, this new findings of G₂, N₆ and N₇ types of trichome in Vitex sp. can be useful in solving placement of problematic members in the family. It is suggested for further studies to identify and establish the species by conducting a research based on their DNA to confirm that either these two unknown V. sp.1 and V. sp.2 are the varieties as they shared the most common trichome characters in Vitex species. If none of the current known species resemble the collected unknown species, it is great to recommend that the species to be viewed as a new member of the genus.

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Author contributions

All authors contributed toward data analysis, drafting and critically revising the paper and agree to be accountable for all aspects of the work.

Disclosure of conflict of interest

The authors have no disclosures to declare.

Compliance with ethical standards

The work is compliant with ethical standards.

References

[1] Stevens, P.F. (2001 onwards). [Online]. [Accessed 3rd November 2014]. Available from World Wide Web: http://www.mobot.org/MOBOT/research/APweb/

[2] Harley, R.M., Atkins, S., Budntsev, P.D., Cantino, P.D., Conn, B.J., Grayer, R., Harley, M.M., de Kok, R., Kresstovskaja, T., Morales, R., Paton, A.j., Ryding, O. & Upson, T. (2004). Labiatae. In

- The Families and Genera of Vascular Plant: Flowering Plant-Dicotyledon. Ed. K. Kubitzki. (Germany: Springer Verlag.), pp. 267–268.
- [3] Corner, E.J.H. (1988). Wayside Tree of Malaya Vol. 1. 3rd edition (Kuala Lumpur: United Selangor Press) pp. 740–756.
- [4] Cantino, P.D., Harley, R. & Steven, W. (1992). Genera of Labiatae: Status and Classification. In *Advances in Labiatae Science*. Ed. R. Harley, (United Kingdom: Royal Botanic Gardens), pp. 511–512.
- [5] LaFrankie, J. (2010). Trees of Tropical Asia: An Illustrated Guide to Diversity (Philippines: Black Tree Publications, Inc.)
- [6] Lam, H.J. (1919). Verbenaceae of the Malay Archipelago (Groningen: Ly. M. DE. Waal) pp. 164 214.
- [7] Ridley, H.N. (1923). Verbenaceae. In *Flora of the Malay Peninsula* (London: Reeve and Co. Ltd.), pp. 2:630–636
- [8] Kochummen, K.M. (1978). Verbenaceae. In F. S. P. Ng (Ed.): Tree Flora of Malaya, 3rd edition (Malaysia: Forest Research Institute Malaysia) pp. 310–313.
- [9] Turner, I.M. (1995). A Catalogue of the vascular plants of Malaya. *The Garden's Bulletin, Singapore*, 47(1), 256-265.
- [10] Bramley, G.L.C. (2019). Flora Malesiana, Series I: Volume 23. Lamiaceae. (National Park Board Singapore.)
- [11] Henderson, M.R. (1959). Malayan Wild Flowers Dicotyledons (Kuala Lumpur: Caxton Press Ltd.)
- [12] Ascensao, L., Nata'lia, M. & Salqme, P.M. (1997). Peltate Glandular Trichomes of *Leonotis leonurus* Leaves: Ultrastructure and Histochemical Characterization of Secretions. *International. Journal of Plant Science*, 158, 249–258.
- [13] Cantino, P.D. (1990). The Phylogenetic Significance of Stomata and Trichomes in the Labiatae and Verbenaceae. *Journal of Arnold Arboretum*, 71, 323–370.
- [14] Xiang, C-L., Dong, Z-H., Peng, H. & Liu, Z-W. (2010). Trichome micromorphology of the East Asiatic genus Chelonopsis (Lamiaceae) and its systematic implications. *Flora*, 205, 434–441.
- [15] Seyedi, Z. & Salmaki, Y. (2015). Trichome morphology and its significance in the systematics of Phlomoides (Lamiaceae; Lamioideae; Phlomideae). *Flora*, 213, 40 48
- [16] Abu-Assab, M.S. & Cantino, P.D. (1987). Phylogenetic Implications of Leaf Anatomy in Subtribe Melittidinae (Labiatae) and Related Taxa. *Journal of Arnold Arboretum*, 68, 1–34.
- [17] Ascensao, L., Mota, L. & Castro, M.D.M. (1999). Glandular Trichomes on the Leaves and flowers of Plectranthus Ornatus: Morphology, Distribution and Histochemistry. *Annals of Botany*, 84, 437–447.

- [18] Metcalfe, C.R. & Chalk, L. (1950). Anatomy of the Dicotyledons. Vol. 2. (London: Oxford Press.)
- [19] Martínez-Natarén, D. A., Parra-Tabla, V., Dzib, G. & Calvo-Irabién, L. M. (2011). Morphology and Density of Glandular Trichomes in Populations of Mexican Oregano (Lippia graveolens H.B.K., Verbenaceae), and the Relationship Between Trichome Density and Climate. *The Journal of the Torrey Botanical Society*, 138(2), 134–144.
- [20] Salmaki, Y., Zarre, Sh., Jamzad, Z. & Br¨auchler, C. (2009). Trichome Micromorphology of Iranian Stachys (Lamiaceae) with Emphasis on its Systematic Implication. *Flora*, 204, 371–381.
- [21] M, Sjana. & P, Sunojkumar. (2018). Trichome Micromorphology and its Systematic Significance in Asian *Leucas* (Lamiaceae). *Flora*, 242, 70-78.
- [22] Celep, F., Kahraman, A., Atalay, Z. & Doğan, M. (2014). Morphology, Anatomy, Palynology, Mericarp and Trichome Micromorphology of the Rediscovered Turkish Endemic *Salvia quezelii* (Lamiaceae) and their taxonomic implications. *Plant Systematics and Evolution*, 300(9), 1945–1958.
- [23] Abbas Azimi, R., Jamzad, Z., Sefidkon, F. & Bakhshi-Khaniki, Gh. (2006). The Potential Value of Phytochemical and Micromorphological Characters in Taxonomic Treatment of Genus *Vitex* L. (Lamiaceae). *The Iranian Journal of Botany*, 12(1), 15-35.
- [24] Henderson, M.R. (1930). Additions to the flora of the Malay Peninsula. *The Garden's Bulletin, Singapore*, 5, 96.
- [25] Steenis, C.G.G.J.Van. (1957). Miscellaneous Botanical Notes VII. Blumea, 15(2), 515–517.
- [26] Henderson, M.R. (1974). *Malayan wild flowers* (Kuala Lumpur: Art Printing works.) pp. 386–387.
- [27] Burkill, I. H. (1935). *A Dictionary of the Economic Products of the Malay Peninsula*. Vol. II 3rd printing 1993. (Kuala Lumpur: Governments of Malaysia and Singapore by the Ministry of Agriculture and Co-operatives, Kuala Lumpur, Malaysia.) pp. 2277–2282.
- [28] Veldkamp, J.F. (1987). Manual for the description of flowering plant. In *Manual of Herbarium Taxonomy*. Ed. E. F. de Vogel. (Jakarta: UNESCO), pp. 20–76.
- [29] Hickey, L.J. (1973). Classification of the Architecture of Dicotyledonous Leaves. *American Journal of Botany*, 60(1), 17-33.
- [30] Ellis, B., Douglas, C.D, Hickey, L.J., John, D.M., Kirk, R.J., Peter, W. & Scott, L.W. (2019). *Manual of Leaf Architecture morphological description and categorization of dicotyledonous and net-veined monocotyledonous angiosperms by Leaf Architecture Working Group.* (Washington D.C.: Cornell University Press).
- [31] Nor Nafizah, M.N. (2006). Comparative Leaf Anatomy, Phytochemistry and Palynology of Nine Genera of Dipterocarpoideae (Dipterocarpaceae), (PhD Thesis, The University of Reading, United Kingdom) pp. 32 45.
- [32] Cutler, D.F., Botha, T. & Stevenson, D.Wm. (2008). *Plant Anatomy an Applied Approach* (Oxford: Blackwell Publishing) pp. 125 250.

- [33] Johansen, D. A. (1940). *Plant Microtechnique*. (Bombay: Tata Mc Graw-Hill Book Company) pp. 16 118.
- [34] Metcalfe, C.R. & Chalk, L. (1972). Verbenaceae. In *Anatomy of the Dicotyledons 2* (Oxford: Oxford University Press.), pp. 1030 –1041
- [35] Payne, W. (1978). A Glossary of Plant Hair Terminology. Brittonia, 30, 239–255.
- [36] Abu-Asab, M.S. & Cantino, P.D., (1994). Systematic Implications of Pollen Morphology in Subfamily Lamioideae and Pogostemonoideae (Labiatae). *Annals of the Missouri Botanical Garden*, 81, 653–686.
- [37] Navarro, T. & El-Oualidi, J. (2000). Trichome Morphology in Teucrium L. (Labiatae). A Taxonomic Review. *Anales del Jardin Botanico de Madrid*, 57, 277–297.
- [38] Wu, C.Y. & Li, H.W. (1965). Materiae Ad Floram Labiatarum Sinensium (1). *Acta Phytotaxonomica Sinica*, 10, 150–154.
- [39] Wu, C.Y. & Li, H.W. (1977). *Chelonopsis. Flora Reipublicae Popularis Sinica*, Vol. 65(2). (Beijing: Science Press) pp. 394–408.
- [40] Nor Nafizah, M.N., Fatimah, M., Norhayati, D., Hasimah, A., Mazatul Azrin, R. & Hilda, E. (2018). Comparative Anatomical Study and Morhology Observation of Three Medicinal *Vitex* Species (Lamiaceae). *Malayan Nature Journal*, 70(3), 285-296
- [41] Celep, F., Kahraman, A., Atalay, Z. & Doğan, M. (2011). Morphology, Anatomy and Trichome Properties of *Lamium truncatum* Boiss. (Lamiaceae) and Their Systematic Implications. *Australian Journal of Crop Science*, 5(2),147-153.